

# The glycopeptide domain of the rat vasopressin precursor

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Received 13 September 1983

The vasopressin precursor is composed of 3 domains, namely vasopressin, MSEL-neurophysin and a glycopeptide. Processing occurs during axonal transport from hypothalamus to neurohypophysis from which the 3 fragments can be isolated. The glycopeptide fragment of the rat vasopressin precursor has been purified and sequenced. Despite the fact that rat MSEL-neurophysin is shortened (93 residues instead of 95 for other mammals), rat glycopeptide has 39 residues, as do the other mammalian glycopeptides, suggesting a similar processing. Fifteen substitutions are however observed when compared to ox glycopeptide. The C-terminal part of MSEL-neurophysin (residues 77–93) and the glycopeptide are encoded by the same exon and the homologies when compared with their bovine counterparts are 58% and 62% respectively. In contrast, the central part of rat MSEL-neurophysin (residues 10–76), which is encoded by a separate exon, displays 96% of homology; vasopressin and the N-terminal part of MSEL-neurophysin (residues 1–9), encoded by a third exon, are nearly invariant.

*Neurohypophysial glycopeptide sequence*  
*Rat glycopeptide*

*Vasopressin-neurophysin precursor*  
*Exon evolution*

## 1. INTRODUCTION

Mammalian vasopressin is a fragment of a 3-domain precursor that is usually processed into vasopressin (9 residues), MSEL-neurophysin (93–95 residues) and a glycopeptide (39 residues) [1–3]. Rat vasopressin has been identified as arginine vasopressin [4], current in most placental mammals. Rat MSEL-neurophysin however is somewhat different from other mammalian MSEL-neurophysins, having 93 residues instead of 95 because of an apparent C-terminal deletion [5]; furthermore, a single arginine is found instead of the penultimate Arg–Arg sequence typical of MSEL-neurophysins [6]. It is therefore of interest to characterize the glycopeptide in order to check whether the processing has been modified by these variations.

## 2. MATERIALS AND METHODS

Freeze-dried posterior pituitary lobes (740 mg,

about 1800 glands) are extracted with 0.1 M HCl (25 mg/ml) and the supernatant is subjected to a molecular sieving on Sephadex G-75 under conditions described [5]. The fraction corresponding to 'crude' neurophysins (50 mg) is collected and chromatographed on a column (0.5 × 38 cm) of DEAE–Sephadex A-50 equilibrated with 0.4 M pyridine acetate pH 5.9 [5]. The glycopeptide filters through the column (8.6 mg) whereas neurophysins, retained, are eluted by an ionic strength gradient [5]. The glycopeptide, adsorbed at pH 7.4 on a column of concanavalin A, appears homogeneous. It is split either with trypsin [5] or with staphylococcal proteinase [7] and resulting peptides are separated by peptide mapping [8]. Peptides are analyzed [10] and amino acid sequences are determined by a manual Edman procedure [10] either directly or after cleavage by subtilisin, isolation of sub-fragments and determination of their sequences. Phenylthiohydantoin amino acids are identified by thin-layer chromatography [11].

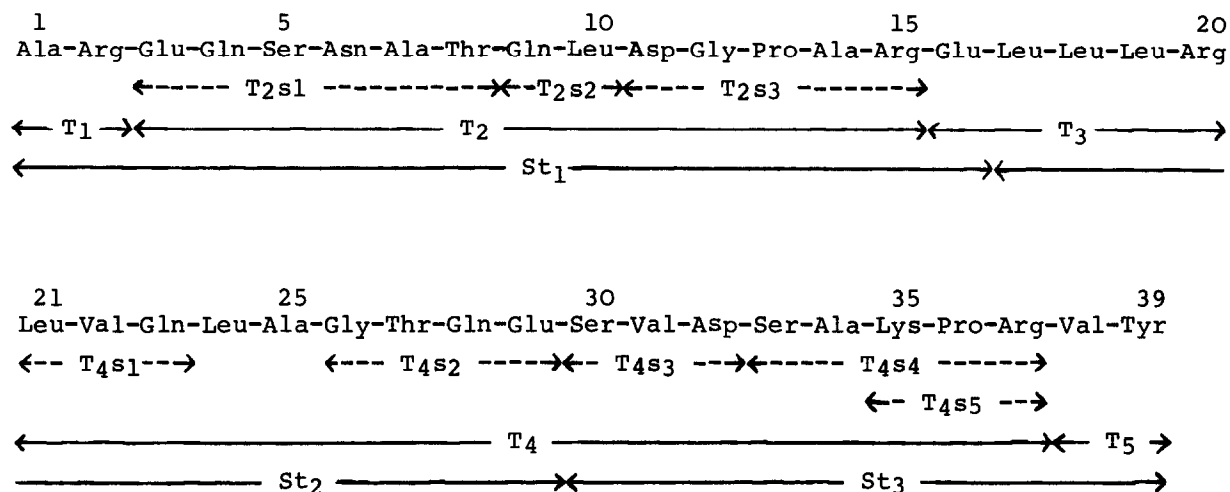
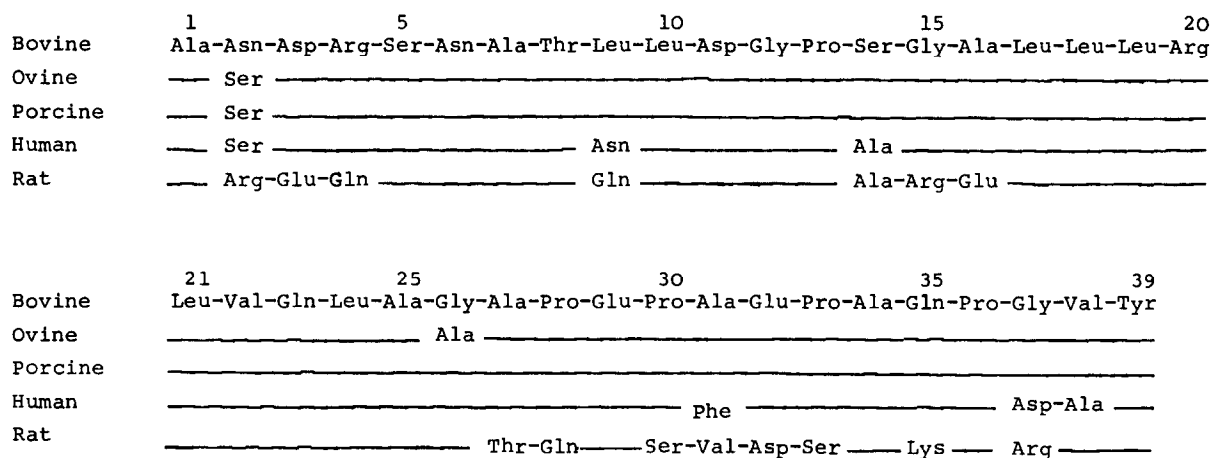


Fig.1. Amino acid sequence of rat glycopeptide. Tryptic peptides (T<sub>1</sub>–T<sub>5</sub>) are sequenced either directly or through subtilisin subfragments (T<sub>2</sub>s<sub>1</sub>, etc....). Overlapping staphylococcal proteinase peptides (St<sub>1</sub> to St<sub>3</sub>) give the alignment of the tryptic peptides.

Table 1  
Amino acid compositions of tryptic peptides of rat glycopeptide

	T <sub>1</sub> (28 nmol)		T <sub>2</sub> (6.3 nmol)		T <sub>3</sub> (3.6 nmol)		T <sub>4</sub> (4.4 nmol)		T <sub>5</sub> (20 nmol)	
	Analysis	Sequence	Analysis	Sequence	Analysis	Sequence	Analysis	Sequence	Analysis	Sequence
Lys							1.00	(1)		
His										
Arg	1.00	(1)	1.00	(1)	1.00	(1)	1.02	(1)		
Asp			1.98	(2)			1.18	(1)		
Thr			0.89	(1)			1.02	(1)		
Ser			1.16	(1)			2.16	(2)		
Glu			2.87	(3)	0.75	(1)	3.23	(3)		
Pro			1.15	(1)			1.00	(1)		
Gly			1.20	(1)			1.34	(1)		
Ala	0.62	(1)	2.28	(2)			2.45	(2)		
1/2 Cys										
Val							2.18	(2)	1.00	(1)
Met										
Ile										
Leu			1.13	(1)	2.60	(3)	1.84	(2)		
Tyr									0.50	(1)
Phe										
CysO <sub>3</sub> H										
Glucosamine <sup>a</sup>			2.91							
Number of residues		2		13		5		17		2
Location in the sequence		1–2		3–15		16–20		21–37		38–39

<sup>a</sup> Uncorrected for destruction



displays a few variations in its first and third parts [6,15], but very rarely in the hormonal part [17]. The substitutions observed in the vasopressin precursor are hardly explained by selective pressure but seem rather relevant to the neutral drift [18].

#### ACKNOWLEDGEMENTS

The authors are grateful to Dr A.F. Parlow of the NIAMD Rat Pituitary Hormones Program for the generous supply of rat posterior pituitaries. They wish to thank Mrs Danielle Thévenet and Miss Christine Jourdain for their skilled technical assistance. This work was supported in part by grants from the CNRS (ERA no.070563), DGRST (no.80-7-294) and the Fondation pour la Recherche Médicale.

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